

Biochemical detection and monitoring of alcohol abuse and abstinence

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SUMMARY. The merits and limitations of traditional and new markers for alcohol abuse (and abstinence) are critically examined for detection and monitoring of alcoholics, hazardous drinkers and binge drinkers. The traditional markers discussed include γ -glutamyltransferase (GGT), aspartate and alanine aminotransaminases (AST, ALT) and mean corpuscular volume (MCV); new markers include mitochondrial AST, carbohydrate-deficient transferrin (CDT), serum/urine 5-hydroxytryptophol, β -hexosaminidase and acetaldehyde adducts. The strengths and weaknesses of several of the self-reporting screening questionnaires are also explored. No laboratory test is reliable enough on its own to support a diagnosis of alcoholism. Sensitivities and specificities vary considerably and depend on the population concerned. GGT continues to remain the test that combines greatest convenience and sensitivity; its diagnostic accuracy can be enhanced by combination with other traditional markers (AST, ALT, MCV). None of the newer markers offers significant advantage, although CDT seems to be better at monitoring patients for increased alcohol consumption or progress towards abstinence.

INTRODUCTION

Alcoholism represents a serious health issue with major socio-economic consequences. Physicians are likely to identify only 20–50% of patients with alcoholism who are attending for medical care.^{1,2} The diagnosis is often based on the patient's self-reporting of alcohol consumption, which is notoriously unreliable^{3,4} and requires a high degree of clinical suspicion. The physician may fail to appreciate the significance of many of the clinical symptoms and findings, which frequently mimic other diseases. Many heavy drinkers are in good health, with no apparent ill-effects from their excessive drinking, and are unlikely to seek medical advice;⁵ they can skilfully hide their addiction and, when medical advice is finally sought, it is often too late to reverse either the organ damage or the dependence on alcohol. However, if alcohol problems are recognized at an early stage, a physician may be able to prevent their further development and progression.^{6–8}

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Recommended safe limits and definitions

The safe limits for alcohol intake are controversial, as there is a wide variation in individual susceptibility to damage. Guidelines drawn up by the Royal College of Physicians advise a weekly limit of 21 units (168 g) of alcohol for men and 14 units (112 g) for women;⁹ these limits have also been endorsed by the Royal Colleges of Psychiatrists and General Practitioners. One unit of alcohol contains 8 g of absolute alcohol. A can of beer or lager (450 mL) contains about 1.5 units of alcohol, a bottle of wine (750 mL) 7–10 units and a bottle of spirits around 30 units.

In 1996, 27% of men and 14% of women in Britain were consistently drinking more than these recommended limits, potentially putting their health at risk ('hazardous drinking').¹⁰ In the age group 16–24 years, 35% of men and 22% of women exceed these amounts.¹¹ Although alcohol abuse can affect any individual in society, personal risk factors include a family history of alcoholism, certain occupations (e.g. publicans, business men who have long absences from family and friends), being a single man and stress.^{11,12}

Definitions of different kinds of drinkers are as follows:

- *Social drinkers* usually drink not more than 2–3 units of alcohol per day and do not become intoxicated. They are unlikely to harm themselves or others through drinking. Alcohol consumption at these levels reduces the risk of developing coronary heart disease by up to 40%.^{13,14}
- *Heavy drinkers* regularly drink more than 6 units a day, but without apparent immediate harm.
- *Problem drinkers* experience physical, psychological, social, family, occupational, financial or legal problems attributable to drinking (but without evidence of alcohol dependence). Consumption of alcohol generally exceeds the recommended safe limits, but may not necessarily be regular. Harmful levels of consumption are > 50 units (400 g) of alcohol per week for men and 35 units (280 g) for women. Estimates suggest that approximately 6% of men and 2% of women exceed these levels.¹¹
- *Alcohol dependence*: people with a compulsion to drink, who take roughly the same amount each day, have increased tolerance in the early stages and reduced tolerance later, suffer withdrawal states if alcohol is reduced or stopped and in whom drinking takes precedence over other activities (pleasures/interests). They persist in using alcohol despite clear evidence of harm. It is difficult to estimate the true number of dependent drinkers in Britain, but recent figures suggest 7.5% of men and 2.1% of women aged 16–64 years.¹⁵

CLINICAL FEATURES OF ACUTE ALCOHOLIC INTOXICATION

The behavioural and clinical features of acute alcoholic intoxication are summarized in Table 1.

TABLE 1. *Clinical features and effects of various alcohol concentrations*

Blood alcohol concentration (mg/100 mL)	Clinical effects
20	Euphoria
30	Increased likelihood of having an accident
40	Disinhibited
80	Impaired coordination, legal limit for driving in UK
150	Loss of self-control, slurred speech, drowsiness, amnesia
300	Stupor, coma
500	Coma, death possible
600	Death certain

Adapted from Refs 9 and 158.

Blood alcohol levels necessary to produce intoxication depend on a number of factors, including the rate of intake (a rapid rise in blood alcohol concentration produces greater effects than a gradual increase), the degree of individual tolerance (which is increased by previous regular alcohol consumption) and whether the individual is drinking in company or in isolation.

PHYSICAL HEALTH HAZARDS ASSOCIATED WITH ALCOHOL ABUSE

The physical health hazards and diseases associated with excessive alcohol consumption are well known (Table 2). Regular consumption of 7.5 units (60 g) of alcohol per day in men and 5 units (40 g) per day in women is associated with increasing risk of any of these diseases. Alcohol abuse can also lead to psychiatric, psychological and social problems and in pregnancy is associated with fetal alcohol syndrome; the Royal College of Obstetricians and Gynaecologists has issued guidelines on this condition (see www.rcog.org.uk/guidelines/alcohol.html; July 2001).

DETECTION OF EXCESSIVE ALCOHOL INTAKE

No exact symptom or clinical finding identifies alcohol abuse in its early stages. Le Go, a French physician, has described subtle physical signs of problem drinking and grouped them in a 'Le Go grid'.^{16,17} The cardinal signs are abnormal skin vascularization (particularly facial telangiectasia), conjunctival injection, coating of the tongue, tremor of the mouth, tongue and hands. The subsequent work of Skinner *et al.*¹⁸ concluded that clinical signs (Le Go grid, tandem gait, deep knee bend, oedema of soft palate, bruises, abrasions, cigarette burns and trauma-related scars) distinguished a group of

TABLE 2. *Disorders and diseases associated with excessive alcohol intake*

Organ or system	Disease or disorder
Liver	Fatty infiltration, alcoholic hepatitis, cirrhosis, liver failure
Gastrointestinal	Oesophagitis, gastritis, Mallory–Weiss tear, haematemesis, peptic ulceration, oesophageal cancer, pancreatitis, diabetes mellitus, diarrhoea, impaired absorption, weight loss, malnutrition
Nervous	Acute intoxication, 'black-outs', epilepsy, tremor (delirium tremens), dementia, Wernicke's encephalopathy, Korsakoff's psychosis, strokes, subdural haematoma (head injury), subarachnoid haemorrhage, hallucinations, peripheral neuropathy
Cardiovascular	Hypertension, arrhythmia, heart failure
Respiratory	Tuberculosis, fractured ribs, pneumonia
Lipids	Hypertriglyceridaemia
Gonads/reproduction	Men: erectile dysfunction, loss of libido, impaired spermatid function (sub- or infertility), small testes, loss of sexual hair Women: menstrual irregularities, subfertility, sexual problems, loss of secondary sexual characteristics
Endocrine	Pseudo-Cushing's, hypoglycaemia
Fetus	Fetal alcohol syndrome

excessive drinkers from social drinkers better than either clinical history or laboratory markers. In contrast, a World Health Organization (WHO) collaborative study found that clinical findings were generally poor as early indicators of alcohol-related harm.¹⁹

Various self-reporting screening questionnaires and laboratory tests ('markers') are now available to assist in the diagnosis of alcoholism. This review briefly describes the strengths and weaknesses of several of the questionnaires that are currently used in screening for excessive drinking and reports on both the conventional and new biochemical markers.

Questionnaires

The ideal questionnaire should be easy to administer, short and accurate, and correlate with other more complex diagnostic procedures and tests. Questionnaires are ideally suited for population screening and can identify up to 80% of alcoholics, but they rely on the patient's truthfulness and memory and are highly susceptible to deliberate concealment. The best known are CAGE, MAST and AUDIT (see below). Both CAGE and MAST are concerned with abnormal drinking behaviour such as drinking in the morning and alcohol-related problems rather than the level of consumption. The more recently developed AUDIT (Alcohol Use Disorders Identification Test) displays high sensitivity and has found increasing use in primary care; it is concerned with the level and frequency of consumption and adverse consequences.

CAGE asks four short questions.²⁰ CAGE is a mnemonic based on the key words in each question: Have you ever felt you should **C**ut down on your drinking? Have people **A**nnoyed you by criticizing your drinking? Have you ever felt **B**ad or **G**uilty about your drinking? Have you ever had a drink first thing in the morning to steady your nerves or to get rid of a hangover ('**E**ye opener')?

Two or more positive answers are generally reliable in diagnosing alcohol dependence, and even one positive answer requires a more detailed clinical assessment. In clinical studies, sensitivities have been 60–95%,^{20–23} specificities 40–95%.^{24,25} A major criticism of CAGE is that a patient may score positive if he/she has religious concerns regarding alcohol, feels guilty even if he/she takes one drink, feels the need to cut down when drinking 'safe' amounts or is very health conscious.

A variant of the CAGE questionnaire is the T-ACE which has been shown to be particularly useful in the detection of heavy alcohol consumption in pregnant women.^{26,27} In T-ACE, questions 2 and 4 are the same as in CAGE and score 1 point each. Questions 1 and 3 are as follows: (1) How many drinks does it take to make you feel high? (Tolerance: the patient is considered tolerant if it takes more than two drinks to make her feel high, 2 points); (3) Have you ever felt you ought to **C**ut down your drinking? (1 point). A total score of ≥ 2 points correctly identifies over 70% of heavy drinkers in pregnancy.

The Michigan Alcoholism Screening Test (MAST)²⁸ asks 25 questions which relate to

recognition of a drinking problem, help-seeking behaviour and alcohol-related disabilities.²⁹ Both sensitivity and specificity have been reported as 85%,³⁰ but other studies have yielded much lower figures than this.^{31,32} There are several modifications of MAST. The Self-Administered Alcoholism Screening Test (SAAST),³³ an improved version of MAST, takes longer to administer and score (35 questions).

Developed by the WHO,³⁴ AUDIT's ten questions deal with quantity and frequency of alcohol consumption, symptoms of early and more established alcohol dependence, and the adverse consequences of drinking. Both sensitivity and specificity are 80–95%.^{34,35}

Laboratory markers

Ideally, laboratory markers should reflect an individual's consumption of alcohol both chronically (screening marker) and acutely (relapse marker). A screening marker should display high sensitivity and specificity and discriminate between safe social drinking and heavy, hazardous drinking. The marker should not be elevated by non-alcohol-induced organ damage (particularly non-alcoholic liver disease) and should be non-invasive (e.g. urine, saliva, breath or blood test). A marker used to detect relapse should be sensitive to any consumption above safe levels. No such marker exists at present.

Laboratory tests are useful in helping to identify those drinking to excess, but they cannot diagnose alcoholism definitively. They can be used only to enhance suspicion and must be combined with a clinical history (including collaborative history from a relative if possible), physical examination, questionnaires and self-reporting. While some studies have shown laboratory markers to be less valuable than physical examination or questionnaires,^{18,19} they do give objective information regarding alcohol consumption and changes in consumption over time, and they help the clinician decide on the possible role of alcohol in a clinical problem or disease process. They are also useful in following up an alcoholic patient and provide motivational input to the patient.

CONVENTIONAL LABORATORY MARKERS

Blood/urine/breath ethanol

Measurements of blood, urine and breath alcohol concentrations have a limited, but

important, role. They provide no information regarding severity of alcohol drinking, but when positive do give objective evidence of recent drinking and can identify increased tolerance. Many heavy drinkers will abstain for 24 h before attending their physician, which results in a very low sensitivity to this test.

Serum γ -glutamyl transferase

Serum γ -glutamyl transferase (GGT) activity is increased in the serum in hepatobiliary disorders and with fairly heavy consumption of alcohol.³⁶ Serum levels of GGT have been found to be elevated in about 75% of individuals who are alcohol-dependent,^{37–39} with a range in sensitivity of 60–90%.^{40–42} The sensitivity is greatest when alcoholics and chronic heavy drinkers are compared to teetotallers and infrequent social drinkers.⁴³ In groups consuming hazardous amounts of alcohol but with no evidence of dependence, the sensitivities are much lower (20–50%), particularly in the primary care setting.^{42,44–47} The increase in serum GGT in response to different amounts and duration of alcohol consumption varies considerably between individuals.⁴⁸ Consumption of > 40 g (> 5 units) of alcohol per day significantly elevates serum GGT in chronic alcoholics, whereas in previous non-drinkers at least 60 g/day for a minimum of 5 weeks is required before any increase occurs. GGT is rarely elevated in subjects under the age of 30 years and is less sensitive in women.⁴⁹

In the general population, progressively higher serum GGT activities are associated with increasing levels of alcohol consumption. Elevated serum GGT is found in 20% of men and 15% of women who consume around 40 g alcohol per day and in 40–50% of men and 30% of women who drink more than 60 g/day. GGT is, then, primarily an indicator of chronic consumption of large amounts of alcohol and is not increased by binge drinking in non-alcohol abusers, unless there is concomitant liver disease. The half-life of GGT is between 14 and 26 days and its level usually returns to normal in 4–5 weeks after drinking ceases.⁵⁰

As well as low sensitivity in some clinical situations, one of the major drawbacks to GGT as a marker of excessive alcohol consumption is its lack of specificity, which can vary from 55 to 100%. Numerous other disorders and drugs can elevate GGT and produce false positive results, including biliary tract disease, non-alcoholic liver disease, obesity, smoking, diabetes mellitus,

pancreatitis, hyperlipidaemia, hyperthyroidism, severe trauma, barbiturates, benzodiazepines, tricyclic antidepressants, anticonvulsants, anticoagulants, inflammation, clotting disorders, cardiac disease and renal disease.⁵¹

Despite its poor specificity, 50–72% of elevated GGT levels can be explained by excessive alcohol consumption.^{40,52} A raised GGT in the absence of the obvious causes listed above should always raise the suspicion of excessive drinking, and a rapid fall in GGT with abstinence is highly suggestive that the initial suspicion was correct. Although GGT is not an ideal screening marker, it is useful in confirming a clinical suspicion of alcoholism and perhaps in monitoring abstinence in the recovering alcoholic.^{53,54}

Serum transaminases

Serum aspartate aminotransferase (AST) and alanine aminotransferase (ALT) are often raised in patients who are alcoholics,^{55,56} although generally not to more than 2–4 times upper normal limits; sensitivities are 25–60% for AST and 15–40% for ALT⁴² (depending on whether the patients are known alcoholics or are being screened for high intake). Serum levels depend markedly on the degree of liver damage and how recently alcohol has been consumed. Acute alcohol intakes of 3–4 g/kg body weight can lead to moderate transient increases in AST in healthy subjects within 24–48 h. The AST:ALT ratio improves the test: a ratio > 1.5 strongly suggests, and a ratio > 2.0 is almost indicative of, alcohol-induced damage to the liver.⁵⁷ One study has shown the AST:ALT ratio to be the best of several markers at distinguishing between alcohol-induced and non-alcoholic liver disease.⁵⁸

Mean corpuscular volume

An increased mean corpuscular volume (MCV) follows chronic heavy drinking and correlates with both the amount and frequency of alcohol ingestion,^{53,59} but it can take at least a month of drinking more than 60 g alcohol daily to raise the MCV above the reference range.⁵⁹ It then takes several months of abstinence for MCV to return to normal,⁶⁰ so MCV has no role in monitoring abstinence or relapse. The actual mechanism by which alcohol causes an increase in MCV appears to include a direct toxic effect of alcohol on red blood cells, folic acid deficiency secondary to alcohol abuse and hepatic damage. Causes of a raised MCV other than excessive alcohol consumption include

vitamin B₁₂ deficiency, folic acid deficiency, hypothyroidism, haemolytic disease with associated reticulocytosis, non-alcoholic liver disease, anticonvulsants, azathioprine, zidovudine, age and smoking.

The main weakness of MCV is its low sensitivity both in hospital environments^{42,61–66} and particularly in primary health care,^{63,67–69} with an overall sensitivity of 40–50%, but its specificity is high (80–90%)^{63,65,69,70} and very few tee-totallers and social drinkers will have elevated MCV values.

Lipids

Although increased high-density lipoprotein cholesterol or triglycerides can raise suspicion of excessive alcohol consumption, neither has sufficient sensitivity and specificity to be of use in diagnosis and monitoring.

Other markers

Serum IgA is classically raised in patients with alcohol-induced liver disease, as are urinary coproporphyrins. Serum urate is also raised in 20–40% of alcoholics. However, none of these markers is useful in screening for alcoholism or monitoring abstinence and all display inadequate sensitivity and specificity.

NEWER MARKERS

Serum mitochondrial AST

Serum AST consists of two isoenzymes: mitochondrial AST (mAST) and cytosolic AST (cAST).⁷¹ In serum samples of normal healthy individuals cAST makes up > 90% of the total activity,⁷¹ but when excessive alcohol consumption selectively injures mitochondria in the liver, mAST is preferentially released.⁷² It is measured by an immunochemical technique in which specific antibodies precipitate cAST, leaving mAST to be measured using standard AST methods.⁷³ A sensitivity of approximately 90% in alcoholic patients has been reported,^{74–76} although the specificity is low (in one study 79% of patients with non-alcoholic liver disease had elevated mAST).⁷⁵ The specificity was improved to 82% by using the mAST:total AST (mAST:tAST) ratio (cut-off 7%),⁷⁵ but with the consequence of lower sensitivity (range 52–100%).^{74–76} The mAST:tAST ratio returns to normal levels within a few weeks of abstinence.

Like most other markers, both mAST and the mAST:tAST ratio have lower sensitivity (30–40%) in screening for hazardous drinking in the community and primary care.^{77–79} The mAST:

tAST ratio can be particularly useful in a liver unit in helping to distinguish alcohol-induced from non-alcohol-related liver damage.

Serum carbohydrate-deficient transferrin

A component of serum transferrin that has an abnormally high isoelectric point – known as desialylated transferrin, later renamed carbohydrate-deficient transferrin (CDT) – was discovered in 1976 in the cerebrospinal fluid of alcoholic patients with cerebellar degeneration⁸⁰ and a few years later in the serum of alcoholic patients.^{81–83} Transferrin exists in normal serum in various forms containing 0–9 sialic acid residues, predominantly tetrasialotransferrin (80%).^{84–86} CDT consists of the asialo, mono-sialo and disialo isoforms, which are less negatively charged at pH 7.4. CDT was first proposed as a marker of excessive alcohol consumption in 1979.⁸⁷

Alcoholic subjects consuming 50–80 g of alcohol per day for at least a week will show increased serum levels.⁸⁸ During abstinence CDT normalizes with a half-life of 15 days^{88,89} and it thus remains elevated for several weeks. If drinking resumes, lower levels of alcohol intake can lead to a rapid re-elevation.⁸⁹ Initial studies assessing the alcohol-induced increase in CDT were conducted in alcoholics and chronic heavy drinkers in detoxification centres; in patients admitted to a general medical department, much more alcohol was required to increase CDT.⁹⁰ Furthermore, in healthy non-alcoholics, CDT failed to rise above the normal reference range after consumption of up to 80 g of alcohol per day for 3 weeks.^{91,92}

CDT can be separated from transferrin by isoelectric focusing or chromatography.^{88,89,93} A popular method of measuring CDT uses a competitive enzyme immunoassay kit (CDTect™) produced by Pharmacia Diagnostics (reference range up to 20 U/L in men, 26 U/L in women) or an Axis %CDT kit (turbidimetric immunoassay), when > 6% CDT:total transferrin is considered elevated.

Stibler's 1991 review⁸⁸ summarizing 20 studies involving 2500 established alcoholics calculated overall sensitivity as 82% and specificity as 97%. Subsequent studies showed that CDT was much better at detecting alcoholics than hazardous drinkers^{63,89,94} and showed sensitivities in less extreme populations (e.g. women, general population, male university students, occasional heavy drinkers, college students, young male soldiers, healthy volunteers) of only

12–45%.^{45,63,69,79,92,94–98} CDT is useless as a screening test for alcohol abuse; a recent meta-analysis of 110 clinical studies showed it to be no better than GGT in this respect.⁹⁹ In an attempt to compensate for the low sensitivity the CDT:total transferrin ratio has been proposed as a better marker.¹⁰⁰

The main strength of CDT is its high specificity – 80–95% in the vast majority of studies^{88,94–97} – but several conditions and diseases reduce this specificity. False positives can occur with: non-alcoholic liver disease (primary biliary cirrhosis,¹⁰¹ chronic active hepatitis,¹⁰¹ chronic hepatitis C¹⁰² and hepatocellular carcinoma¹⁰³); the genetic variant of transferrin known as D1 (found in 1% of the black population and < 1% in the white US population¹⁰⁴ and in up to 2% of Northern Europeans¹⁰⁵); and the rare carbohydrate-deficient glycoprotein syndrome.^{88,93}

One study has shown that patients who have undergone combined transplantation of the pancreas and kidneys had elevated CDT concentrations in the absence of alcohol consumption,¹⁰⁶ and increased CDT has also been seen in patients with cystic fibrosis due to defective sialylation.¹⁰⁷ Fagerberg *et al.* found that CDT levels may be altered by insulin-related metabolism (insulin sensitivity): male patients with hypertension who had elevated CDT concentrations had low blood glucose and serum triglyceride levels.^{95,108} Other conditions associated with false positives are iron deficiency,¹⁰⁹ untreated galactosaemia,¹¹⁰ rectal carcinoma, senile dementia, depression, pregnancy and solvent abuse.

In conclusion, CDT is of most use in monitoring patients for an increase in alcohol consumption or progressive abstinence.^{111,112} A systematic review using the results of six prospective outcome studies with 239 male alcoholics found CDT to be significantly more sensitive than GGT in the detection of relapses.¹¹³ It is of no use in screening for heavy alcohol consumption in the general population, particularly in women, and will not identify binge drinkers. However, a review of eight studies comprising 2214 patients found CDT to be better than GGT at distinguishing alcoholic from non-alcoholic liver disease.¹¹³

Serum/urine 5-hydroxytryptophan

Serotonin is normally metabolized to 5-hydroxytryptophol-3-acetic acid (5-HIAA) and 5-hydroxytryptophol (5-HTOL), with 5-HIAA being the major metabolite. Alcohol dose-dependently shifts serotonin metabolism towards 5-HTOL.¹¹⁴

The increase in 5-HTOL and decrease in 5-HIAA can be measured in both blood and urine using either gas chromatography or high-performance liquid chromatography (HPLC).^{114,115} The 5-HTOL:5-HIAA ratio has been found to reflect alcohol intake in the past 24 h^{115,116} and to remain elevated for 6–15 h after blood alcohol has returned to normal.¹¹⁴ This potential marker has high sensitivity (as little as 20 g/day alcohol can be detected)¹¹⁵ and specificity in detecting very recent alcohol consumption, and would be useful where frequent follow-up of patients is feasible and necessary. However, its method of measurement ensures that it will not be routinely used in the majority of laboratories.

Serum β -hexosaminidase

β -Hexosaminidase (β -HEX) is an acid lysosomal glycosidase. Increased serum and urine levels have been reported in alcoholic patients and in healthy volunteers after consumption of > 60 g of alcohol per day for at least 10 days, with sensitivities of 70–90%;^{117–120} this is better than GGT and other established markers. However, like CDT, β -HEX appears not to be as effective in identifying less excessive but still harmful levels of drinking in unselected populations.¹²¹ In alcoholics, β -HEX levels fall rapidly (7–10 days) to normal following abstinence. The β -HEX B isoform in particular is highly indicative of alcohol abuse.¹²² Although high specificities (approximately 90%) have been reported for β -HEX,^{44,120,122} serum levels of β -HEX have been noted to be increased in hypertension, diabetes mellitus, cirrhosis, pregnancy, in users of the oral contraceptive pill, cerebral infarction and myocardial infarction.^{123–125} One of the major potential strengths of β -HEX is that it can be measured using standard and inexpensive laboratory techniques (spectrophotometry and fluorimetry). Its major isoenzymes can also be easily measured; isoform B is heat-stable whereas isoform A is heat-sensitive.¹²⁶

Thus, serum β -HEX is a sensitive, easily measured, inexpensive test for excessive alcohol consumption, but like CDT it does not perform well in unselected populations; moreover, there are conditions other than alcohol intake that may cause it to be elevated.

Serum acetaldehyde and acetaldehyde adducts

Acetaldehyde, the first metabolite of ethanol, is not a good marker of alcohol consumption as it is metabolized to acetate within a few hours of consumption.¹²⁷ Acetaldehyde readily forms

Schiff bases with amines and after irreversible rearrangement forms an acetaldehyde-protein adduct.¹²⁸

Several proteins form adducts with acetaldehyde, including albumin and haemoglobin;^{42,129,130} these can be measured either using HPLC techniques¹³¹ or by detection of new epitopes created from the acetaldehyde-protein adduct using immunoassay.^{132,133} Whole-blood-associated acetaldehyde (WBAA) includes both albumin and haemoglobin adducts, the concentration of the latter being 5–10 times higher than that of adducts with plasma proteins.^{134,135} WBAA is affected by both acute and antecedent drinking behaviour; it returns to normal levels within 3 weeks of abstinence.^{134,135} One drawback of this marker is that acetaldehyde can form in samples artefactually in the absence of alcohol intake.¹³⁶

The haemoglobin-acetaldehyde adduct (HbA1-AcH) is detected in patients with excessive alcohol consumption with sensitivity 25–50%.^{42,133,137,138} In patients admitted to a drug and alcohol rehabilitation unit, sensitivity was reported at 67% and specificity at 77%, better than GGT, AST or MCV; of these markers, HbA1-AcH was the only one significantly correlated with reported alcohol intake and with the theoretical ability to detect heavy hazardous drinking before the onset of liver damage.¹³⁹ Immunoglobulin A (IgA) reactivity with acetaldehyde-modified proteins has been shown to be elevated in both alcoholics and heavy drinkers, but not in social drinkers or patients with non-alcoholic liver disease.^{140,141} However, in pregnant women who had abused alcohol, HbA1-AcH was inferior to both MCV and GGT in detecting excessive alcohol consumption or the adverse effects of alcohol on the fetus.¹⁴²

Other markers

Other potential markers of excessive alcohol intake include fatty acid ethyl esters,¹⁴³ phosphatidylethanol,¹⁴⁴ sialic acid,¹⁴⁵ erythrocyte aldehyde dehydrogenase,¹⁴⁶ plasma α -aminobutyric acid: leucine ratio,¹⁴⁷ urinary salsolinol¹⁴⁸ and urinary dolichols.¹⁴⁹ Many of these require complex measurement techniques outside the capacity of the routine laboratory. All require further clinical evaluation, and none so far offers significant advantages over the existing established markers.

Combinations of markers

Attempts have been made to improve the sensitivity of single laboratory markers by

combining them, but although some of the combinations have shown enhanced sensitivity (e.g. CDT plus GGT, CDT plus MCV),^{58,89,150-155} none has been widely accepted. Sophisticated mathematical treatment of results from multiple laboratory tests has also been proposed,^{154,156,157} but the large number of test parameters required make the approach impractical, and in any case increased sensitivity invariably decreases specificity. Use of two or three different established markers appears to be optimal.

The Driver and Vehicle Licensing Agency (DVLA) has issued guidelines in connection with laboratory markers of excessive alcohol consumption (www.dvla.gov.uk/at_a_glance/ch5_drug_alcohol.htm, July 2001). Laboratory markers are primarily used here when a high-risk offender is being considered for the return of a suspended driving licence, to support a clinical history of either abstinence or controlled 'social' drinking. The Expert Medical Panel which advises the DVLA considers the most convenient markers to be the established ones of GGT, AST and MCV.

CONCLUSIONS

No laboratory test is reliable enough on its own to support a diagnosis of alcoholism; laboratory tests need to be part of a diagnostic process that includes a detailed clinical history and examination and the use of questionnaires. Laboratory markers are useful in both raising the suspicion and confirming the diagnosis of alcohol abuse; they are also helpful in the follow-up of patients undergoing treatment and in monitoring abstinence. However, sensitivities and specificities of the different laboratory markers vary considerably and depend on the population concerned. None has high accuracy in unselected populations in primary care settings and all are weak in screening for harmful heavy consumption of alcohol. The conventional marker GGT continues to remain the test combining greatest convenience and sensitivity. Its diagnostic accuracy can be enhanced by combination with other traditional markers such as AST, ALT and MCV. None of the newer markers offers significant advantage, although CDT seems to be better at monitoring patients for increased alcohol consumption or progress towards abstinence and possibly in distinguishing alcoholic liver disease from non-alcoholic liver disease.

REFERENCES

- Solomon J, Vanga N, Morgan JP, Joseph P. Emergency room physicians: recognition of alcohol misuse. *J Stud Alcohol* 1980; **41**: 583-6
- Persson J, Magnusson PH. Comparison between different methods of detecting patients with excessive consumption of alcohol. *Acta Med Scand* 1988; **223**: 101-9
- Popham RE, Schmidt W. Words and deeds: the validity of self-report data on alcohol consumption. *J Stud Alcohol* 1981; **42**: 355-68
- Watson CG, Tilleskjoer C, Hoedeccheck-Schow EA, Pucel J, Jacobs L. Do alcoholics give valid self-reports? *J Stud Alcohol* 1984; **45**: 344-84
- Anderson P, Cremona A, Wallace P. What are safe levels of alcohol consumption? *BMJ* 1984; **289**: 1657-8
- Bien TH, Miller WR, Tonigan JS. Brief interventions for alcohol problems: a review. *Addiction* 1993; **88**: 315-36
- Bradley KA. Management of alcoholism in the primary care setting. *West J Med* 1992; **156**: 273-7
- WHO Brief Intervention Study Group. A cross-national trial of brief interventions with heavy drinkers. *Am J Public Health* 1996; **86**: 945-55
- Royal College of Physicians. *A Great and Growing Evil - The Medical Consequences of Alcohol Abuse. Report of a Working Party*. London: Tavistock, 1987
- Austoker J. Reducing alcohol intake. *BMJ* 1994; **308**: 1549-52
- Office for National Statistics. Social Survey Division. *Living in Britain. Results from the 1996 General Household Survey*. London: The Stationery Office, 1998
- Gorman DM. Alcohol misuse and the predisposing environment. *Br Med Bull* 1994; **50**: 36-49
- Rimm EB, Giovannucci FL, Willett WC, Colditz GA, Ascherio A, Rosner B, et al. Prospective study of alcohol consumption and risk of coronary heart disease in men. *Lancet* 1991; **338**: 464-8
- Friedman LA, Kimball AW. Coronary heart disease mortality and alcohol consumption in Framingham. *Am J Epidemiol* 1986; **124**: 481-9
- Office for National Statistics. Department of Health. *Statistics on Alcohol: 1976 Onwards*. London: The Stationery Office, 1999
- Le Go PM. Le dépistage précoce de l'alcoolisme. *Presse Méd* 1968; **76**: 578-80
- Le Go PM. *Le Dépistage Précoce et Systématique du Buveur Excessif*. Paris: Département d'Alcologie Thérapeutique de Rion-Laboratories, 1976
- Skinner HA, Holt S, Sheu WJ, Israel Y. Clinical versus laboratory detection of alcohol abuse: the Alcohol Clinical Index. *BMJ* 1986; **292**: 1703-8
- Saunders JB, Aasland OG. *WHO Collaborative Project on Identification and Treatment of Persons with Harmful Alcohol Consumption. Report on Phase 1. Development of a Screening Instrument*. MNH/DAT/86.3. Geneva: WHO, 1987
- Ewing JA. Detecting alcoholism: the CAGE questionnaire. *JAMA* 1984; **252**: 1905-7

- 21 Bernadt MW, Mumford J, Taylor C, Smith B, Murray RM. Comparison of questionnaire and laboratory tests in the detection of excessive drinking and alcoholism. *Lancet* 1982; **i**: 325–8
- 22 King M. At-risk drinking among general practice attenders – validation of the CAGE questionnaire. *Psychol Med* 1986; **16**: 213–7
- 23 Bartu A, Baldwin C, Goonewardene R. *Computerised Screening and Interventions for Problem Drinkers in a Non-Clinical Setting*. Mt Lawley WA: West Australian Alcohol and Drug Authority, 1991
- 24 Simon DG, Eley JW, Greenberg RS, Newman N, Gillespie T, Moore M. A survey of alcohol use in an inner-city ambulatory care setting. *J Gen Intern Med* 1991; **6**: 295–8
- 25 Wallace P, Cutler S, Haines A. Randomised controlled trial of general practitioner intervention in patients with excess alcohol consumption. *BMJ* 1988; **297**: 663–8
- 26 Sokol RJ, Martier SS, Ager JW. The T-ACE questions: practical prenatal detection of risk drinking. *Am J Obstet Gynecol* 1989; **160**: 863–70
- 27 Maisto SA, Connors GJ, Allen JP. Contrasting self-report screens for alcohol problems. A review. *Alcohol Clin Exp Res* 1995; **19**: 1510–6
- 28 Selzer ML. The Michigan Alcoholism Screening Test: the quest for a new diagnostic instrument. *Am J Psychiatry* 1971; **127**: 1653–8
- 29 Crook GM, Oei TPS, Young RM. Structure of the MAST with an Australian sample of alcoholics. *Drug Alcohol Rev* 1994; **13**: 41–6
- 30 Storgaard H, Nielson SD, Gluud C. The validity of the Michigan Alcoholism Screening Test (MAST). *Alcohol Alcohol* 1994; **29**: 493–502
- 31 Saunders WM, Kershaw PW. Screening tests for alcoholism – findings from a community study. *Br J Addiction* 1980; **75**: 37–41
- 32 McIntyre D. Alcohol-related problems among male patients admitted to a general medical ward – their identification and follow-up. *Health Bull* 1979; **37**: 213–7
- 33 Swenson WM, Morse RM. The use of a self-administered alcoholism screening test (SAAST) in a medical center. *Mayo Clin Proc* 1975; **50**: 204–8
- 34 Saunders JB, Aasland OG, Babor TF, De la Fuente JR, Grant M. Development of the Alcohol Use Disorders Identification Test (AUDIT): WHO collaborative project on early detection of persons with harmful alcohol consumption II. *Addiction* 1993; **88**: 791–804
- 35 Allen JP, Litten RZ, Fertig JB, Babor T. A review of research on the Alcohol Use Disorders Identification test (AUDIT). *Alcohol Clin Exp Res* 1997; **21**: 613–9
- 36 Penn R, Worthington DJ. Is serum γ -glutamyl-transferase a misleading test? *BMJ* 1983; **286**: 531–5
- 37 Wu A, Slavin G, Levi AJ. Elevated serum gamma-glutamyl transferase (transpeptidase) and histological liver damage in alcoholism. *Am J Gastroenterol* 1976; **65**: 318–23
- 38 Rosalki SB, Rau D. Serum gamma-glutamyl transpeptidase activity in alcoholism. *Clin Chim Acta* 1972; **39**: 41–7
- 39 Stetter F, Gaertner HJ, Wiatr G, Mann K, Breyer-Pfaff U. Urinary dolichol – a doubtful marker of alcoholism. *Alcohol Clin Exp Res* 1991; **15**: 938–41
- 40 Behrens UJ, Worner TM, Braly LF, Schaffner F, Lieber CS. Carbohydrate-deficient transferrin, a marker for chronic alcohol consumption in different ethnic populations. *Alcohol Clin Exp Res* 1988; **12**: 427–32
- 41 Schellenberg F, Benard JY, Le Goff AM, Bourdin C, Weill J. Evaluation of carbohydrate deficient transferrin compared with Tf index and other markers of alcohol abuse. *Alcohol Clin Exp Res* 1989; **13**: 605–10
- 42 Sillanaukee P, Seppa K, Koivula T, Israel Y, Niemela O. Acetaldehyde-modified haemoglobin as a marker of alcohol consumption: comparison of two new methods. *J Lab Clin Med* 1992; **120**: 42–7
- 43 Conigrave KM, Saunders JB, Whitfield JB. Diagnostic tests for alcohol consumption. *Alcohol Alcohol* 1995; **30**: 13–26
- 44 Karkkainen P, Poikolainen K, Salaspuro M. Serum β -hexosaminidase as a marker of heavy drinking. *Alcohol Clin Exp Res* 1990; **14**: 187–90
- 45 Poupon RE, Schellenberg F, Nalpas B, Weill J. Assessment of the transferrin index in screening heavy drinkers from a general practice. *Alcohol Clin Exp Res* 1989; **13**: 549–53
- 46 Kristenson H. Methods of intervention to modify drinking-patterns in heavy drinkers. In: Galanter M, ed. *Recent Developments in Alcoholism*, Vol. 5. New York: Plenum, 1987: 403–23
- 47 Nilssen O, Forde OH. The Tromsø Study: the positive predictive value of gamma-glutamyl transferase and an alcohol questionnaire in the detection of early stage risk drinkers. *J Int Med* 1991; **229**: 497–500
- 48 Belfrage P, Berg B, Cronholm T, Elmqvist D, Hagerstrand I, Johansson B, *et al.* Prolonged administration of ethanol to young, healthy volunteers: effects on biochemical, morphological and neurophysiological parameters. *Acta Med Scand Suppl* 1973; **552**: 1–44
- 49 Whitfield JB, Hensley WJ, Bryden D, Gallagher H. Effects of age and sex on biochemical responses to drinking habits. *Med J Aust* 1978; **2**: 629–32
- 50 Orrego H, Black JE, Israel Y. Relationship between gamma glutamyl transpeptidase and mean urinary alcohol levels in alcoholics while drinking and after alcohol withdrawal. *Alcohol Clin Exp Res* 1985; **9**: 10–13
- 51 Rosman AS, Lieber CS. Biochemical markers of alcohol consumption. *Alcohol Health Res World* 1990; **14**: 210–8
- 52 Kristenson H, Trelle E, Fex G, Hood B. Serum gamma glutamyl transferase: statistical distribution in a middle aged male population and evaluation of alcohol habits in individuals with elevated levels. *Prev Med* 1980; **9**: 108–19
- 53 Irwin M, Baird S, Smith T, Schuckit M. Use of laboratory tests to monitor heavy drinking by alcoholic men discharged from a treatment program. *Am J Psychiatry* 1988; **145**: 595–9

- 54 Irwin M. Monitoring heavy drinkers in recovering alcoholics. *Alcohol Clin Exp Res* 1987; **11**: 202
- 55 Chan AW, Welte JW, Whitney RB. Identification of alcoholism in young adults by blood chemistries. *Alcohol* 1987; **4**: 175-9
- 56 Rosman AS, Lieber CS. Biological markers of alcoholism. In: Lieber CS, ed. *Medical and Nutritional Complications of Alcoholism*. New York: Plenum, 1992
- 57 Cohen JA, Kaplan MM. The SPOT/SGOT ratio: an indicator of alcoholic liver disease. *Dig Dis Sci* 1979; **24**: 835-8
- 58 Sharpe PC, McBride R, Archbold GPR. Biochemical markers of alcohol abuse. *Q J Med* 1996; **89**: 137-44
- 59 Whitehead TP, Clarke CA, Whitfield AG. Biochemical and haematological markers of alcohol intake. *Lancet* 1978; **i**: 978-81
- 60 Morgan MY, Camil ME, Luck W, Sherlock S, Hoffbrand AV. Macrocytosis in alcohol-related liver disease: its value for screening. *Clin Lab Haematol* 1981; **3**: 35-44
- 61 Behrens UJ, Worner TM, Lieber CS. Changes in carbohydrate deficient transferrin levels after alcohol withdrawal. *Alcohol Clin Exp Res* 1988; **12**: 539-44
- 62 Gjerde H, Johnsen J, Bjorneboe A, Bjorneboe GE, Morland J. A comparison of serum carbohydrate deficient transferrin with other biological markers of excessive drinking. *Scand J Clin Lab Invest* 1988; **48**: 1-6
- 63 Sillanaukee P, Seppa K, Lof K, Koivula T. CDT by anion exchange chromatography followed by RIA as a marker of heavy drinking among men. *Alcohol Clin Exp Res* 1993; **17**: 230-3
- 64 Wickramasinghe SN, Corridan SB, Hasan, Marjot DH. Correlations between acetaldehyde-modified haemoglobin, carbohydrate-deficient transferrin (CDT) and haematological abnormalities in chronic alcoholism. *Alcohol Alcohol* 1994; **29**: 415-23
- 65 Van Pelt J. Carbohydrate-deficient transferrin: a new biochemical marker for chronic excessive alcohol consumption. *Ned Tijdschr Geneesk* 1997; **141**: 773-7
- 66 Aithal GP, Thornes H, Dwarakanath AD, Tanner AR. Measurement of carbohydrate-deficient transferrin (CDT) in a general medical clinic: is this test useful in assessing alcohol consumption. *Alcohol Alcohol* 1998; **33**: 304-9
- 67 Baxter S, Fink R, Leader AR, Rosalki SB. Laboratory tests for excessive alcohol consumption evaluated in general practice. *Alcohol Alcohol* 1980; **15**: 164-6
- 68 Sillanaukee P, Aalto M, Seppa K. Carbohydrate-deficient transferrin and conventional alcohol markers as indicators of brief intervention among heavy drinkers in primary health care. *Alcohol Clin Exp Res* 1998; **22**: 892-6
- 69 Bisson JI, Milford-Ward A. A comparison of carbohydrate deficient transferrin with other markers of alcohol misuse in male soldiers under the age of thirty. *Alcohol Alcohol* 1994; **29**: 315-21
- 70 Skinner HA, Holt S, Schuller R, Roy J, Israel Y. Identification of alcohol abuse using laboratory tests and a history of trauma. *Ann Intern Med* 1984; **101**: 847-51
- 71 Rej R. Aspartate aminotransferase activity and isoenzyme proportion in human liver tissues. *Clin Chem* 1978; **24**: 1971-9
- 72 Ishii H, Okuno F, Shigeta Y, Tsuchiya M. Enhanced serum glutamic oxaloacetic transaminase activity of mitochondrial origin in chronic alcoholics. *Curr Alcohol* 1979; **5**: 101-8
- 73 Rej R. A immunochemical procedure for determination of mitochondrial aspartate aminotransferase in human serum. *Clin Chem* 1980; **26**: 1694-700
- 74 Nalpas B, Vassault A, Le Guillou A, Lesgourgues B, Ferry N, Lacour N, et al. Serum activity of mitochondrial aspartate aminotransferase: a sensitive marker of alcoholism with and without alcoholic hepatitis. *Hepatology* 1984; **4**: 893-6
- 75 Nalpas B, Vassault A, Charpin S, Lacour B, Berthelot P. Serum mitochondrial aspartate aminotransferase as a marker of chronic alcoholism: diagnostic value and interpretation in a liver unit. *Hepatology* 1986; **6**: 608-14
- 76 Okuno F, Ishii H, Kashiwazaki K, Takagi S, Shigeta Y, Arai M, et al. Increase in mitochondrial GOT (m-GOT) activity after chronic alcohol consumption: clinical and experimental observations. *Alcohol* 1988; **5**: 49-53
- 77 Schiele F, Artur Y, Varasteh A, Wellman M, Siest G. Serum mitochondrial aspartate aminotransferase activity: not useful as a marker of excessive alcohol consumption in an unselected population. *Clin Chem* 1989; **35**: 926-30
- 78 Nalpas R, Poupon RE, Vassault A, Hauzanneau P, Sage Y, Schellenberg F, et al. Evaluation of mAST/tAST ratio as a marker of alcohol misuse in a non-selected population. *Alcohol Alcohol* 1989; **24**: 415-9
- 79 Nilssen O, Huseby NE, Hoyer G, Brenn T, Schirmer H, Forde OH. New alcohol markers - how useful are they in population studies? The Svalbard Study 1988-1989. *Alcohol Clin Exp Res* 1992; **16**: 82-6
- 80 Stibler H, Kjellin KG. Isoelectric focusing and electrophoresis of the CSF proteins in tremor of different origin. *J Neurol Sci* 1976; **30**: 269-85
- 81 Stibler H, Allgulander C, Borg S, Kjellin KG. Abnormal microheterogeneity of transferrin in serum and cerebrospinal fluid in alcoholism. *Acta Med Scand* 1978; **204**: 49-56
- 82 Stibler H, Borg S. Evidence of a reduced sialic acid content in serum transferrin in male alcoholics. *Alcohol Clin Exp Res* 1981; **5**: 545-9
- 83 Stibler H, Borg S, Joustra M. Micro anion exchange chromatography of carbohydrate deficient transferrin in serum in relation to alcohol consumption (Swedish patent 8400587-5). *Alcohol Clin Exp Res* 1986; **10**: 535-44
- 84 Schacter H. Biosynthetic controls that determine the branching and microheterogeneity of protein-bound oligosaccharides. *Biochem Cell Biol* 1986; **64**: 163-81

- 85 Van Eijk HG, van Noort WL, Dubelaar M-L, van der Heul C. The microheterogeneity of human transferrin in biological fluids. *Clin Chim Acta* 1983; **132**: 167–71
- 86 Marz L, Hatton M, Berry L, Regoeczi E. The structural heterogeneity of the carbohydrate moiety of desialylated human transferrin. *Can J Biochem* 1982; **60**: 624–30
- 87 Stibler H, Borg S, Allgulander C. Clinical significance of abnormal heterogeneity of transferrin in relation to alcohol consumption. *Acta Med Scand* 1979; **206**: 275–81
- 88 Stibler H. Carbohydrate-deficient transferrin in serum: a new marker of potentially harmful alcohol consumption reviewed. *Clin Chem* 1991; **37**: 2029–37
- 89 Allen JP, Litten RZ, Anton RF, Cross GM. Carbohydrate-deficient transferrin as a measure of immoderate drinking: remaining issues. *Alcohol Clin Exp Res* 1994; **18**: 799–812
- 90 Bell H, Tallaksen CME, Try K, Haug E. Carbohydrate-deficient transferrin and other markers of high alcohol consumption: a study of 502 patients admitted consecutively to a medical department. *Alcohol Clin Exp Res* 1994; **18**: 1103–8
- 91 Lesch OM, Walter H, Antral J, Heggli DE, Kovacz A, Leitner A, et al. Carbohydrate-deficient transferrin as a marker of alcohol consumption: a study with healthy subjects. *Alcohol Alcohol* 1996; **31**: 265–71
- 92 Salmela KS, Laitinen K, Nystrom M, Salaspuro M. Carbohydrate-deficient transferrin during 3 weeks heavy alcohol consumption. *Alcohol Clin Exp Res* 1994; **18**: 228–30
- 93 Bean P. Carbohydrate deficient transferrin in the diagnosis of alcohol abuse: diagnostic performance and clinical significance. *Lab Medica Int* 1997; **14**: 16–9
- 94 Gronbaek M, Henriksen JH, Becker U. Carbohydrate-deficient transferrin: a valid marker of alcoholism in population studies? Results from the Copenhagen City Heart Study. *Alcohol Clin Exp Res* 1995; **19**: 457–61
- 95 Fagerberg B, Agewall S, Berglund A, Wysocki M, Lundberg PA, Lindstedt G. Is carbohydrate-deficient transferrin in serum useful for detecting alcohol consumption in hypertensive patients? *Clin Chem* 1994; **40**: 2057–63
- 96 La Grange L, Anton RF, Crow H, Garcia S. A correlational study of carbohydrate-deficient transferrin values and alcohol consumption among Hispanic college students. *Alcohol Clin Exp Res* 1994; **18**: 653–6
- 97 Nystrom M, Perasalo J, Salaspuro M. Carbohydrate-deficient transferrin (CDT) in serum as a possible indicator of heavy alcohol drinking in young university students. *Alcohol Clin Exp Res* 1992; **16**: 93–7
- 98 Vermes I, van den Berg FA. Clinical utility of carbohydrate deficient transferrin to detect alcohol abuse in a general population. *Clin Chem* 1996; **42**: 2048–50
- 99 Scouller K, Conigrave KM, Macaskill P, Irwig L, Whitfield JB. Should we use carbohydrate-deficient transferrin instead of γ -glutamyltransferase for detecting problem drinkers? A systematic review and metaanalysis. *Clin Chem* 2000; **46**: 1894–902
- 100 Halm U, Tannapfel A, Mossner J, Berr F. Relative versus absolute carbohydrate transferrin as a marker of alcohol consumption in patients with acute alcoholic hepatitis. *Alcohol Clin Exp Res* 1999; **23**: 1614–8
- 101 Rublo M, Caballeria J, Deulofeu R, Caballeria L, Gasso M, Pares A, et al. Carbohydrate-deficient transferrin as a marker of alcohol consumption in male patients with liver disease. *Alcohol Clin Exp Res* 1997; **21**: 923–7
- 102 Stauber RE, Stepan V, Trauner M, Wilderstruschnig M, Leb G, Krejs GJ. Evaluation of carbohydrate-deficient transferrin for detection of alcohol abuse in patients with liver dysfunction. *Alcohol Alcohol* 1995; **30**: 171–6
- 103 Murawaki Y, Sugisaki H, Yuasa I, Kawasaki H. Serum carbohydrate deficient transferrin in patients with nonalcoholic liver disease and with hepatocellular carcinoma. *Clin Chim Acta* 1997; **259**: 97–108
- 104 Bean P, Peter J. Allelic D variants of transferrin in evaluation of alcohol abuse: differential diagnosis by isoelectric focusing-immunoblotting-laser densitometry. *Clin Chem* 1994; **40**: 2078–83
- 105 Giblet ER. The plasma transferrins. *Prog Med Genet* 1962; **2**: 34–63
- 106 Arndt T, Hackler R, Muller T, Kleine TO, Gressner AM. Increased serum concentration of carbohydrate-deficient transferrin in patients with combined pancreas and kidney transplantation. *Clin Chem* 1997; **42**: 344–51
- 107 Larsson A, Flodin M, Kollberg H. Increased serum concentrations of carbohydrate-deficient transferrin (CDT) in patients with cystic fibrosis. *Ups J Med Sci* 1998; **103**: 231–6
- 108 Fagerberg B, Agewall S, Urbanavicius V, Attval S, Lundberg PA, Lindstedt G. Carbohydrate-deficient transferrin is associated with insulin sensitivity in hypertensive men. *J Clin Endocrinol Metab* 1994; **79**: 712–5
- 109 De Feo TM, Fargion S, Duca L, Mattioli M, Cappellini MD, Sampietro M, et al. Carbohydrate-deficient transferrin, a sensitive marker of chronic alcohol abuse, is highly influenced by body iron. *Hepatology* 1999; **29**: 658–63
- 110 Stibler H, von Döbeln U, Kristiansson B, Gutenberg C. Carbohydrate-deficient transferrin in galactosaemia. *Acta Paediatr* 1997; **86**: 1377–8
- 111 Huseby NE, Bjordal E, Nilssen O, Barth T. Utility of biological markers during outpatient treatment of alcohol-dependent subjects: carbohydrate-deficient transferrin responds to moderate changes in alcohol consumption. *Alcohol Clin Exp Res* 1997; **21**: 1343–6
- 112 Reynaud M, Hourcade F, Planche F, Albuissou E, Meunier MN, Planche R. Usefulness of carbohydrate-deficient transferrin in alcoholic patients with normal γ -glutamyltransferase. *Alcohol Clin Exp Res* 1998; **22**: 615–8

- 113 Salaspuro M. Carbohydrate-deficient transferrin as compared to other markers of alcoholism: a systematic review. *Alcohol* 1999; **19**: 261–71
- 114 Helander A, Beck O, Jones AW. Laboratory testing for recent alcohol consumption: comparison of ethanol, methanol and 5-hydroxytryptophol. *Clin Chem* 1996; **42**: 618–24
- 115 Voltaire A, Beck O, Borg S. Urinary 5-hydroxytryptophol: a possible marker of recent alcohol consumption. *Alcohol Clin Exp Res* 1992; **16**: 281–5
- 116 Voltaire Carlsson A, Hiltunen AJ, Beck O, Stibler H, Borg S. Detection of relapses in alcohol-dependent patients: comparison of carbohydrate deficient transferrin in serum, 5-hydroxytryptophol in urine and self-reports. *Alcohol Clin Exp Res* 1993; **17**: 703–8
- 117 Hultberg B, Isaksson A, Tidestrom G. Hexosaminidase, leucine aminopeptidase, cystidyl aminopeptidase, hepatic enzymes and bilirubin in serum of chronic alcoholics with acute alcohol intoxication. *Clin Chim Acta* 1980; **105**: 317–23
- 118 Hultberg B, Isaksson A, Berglund M, Moberg A-L. Serum β -hexosaminidase isoenzyme: a sensitive marker for alcohol abuse. *Alcohol Clin Exp Res* 1991; **15**: 549–52
- 119 Karkkainen P, Jokelainen K, Roine R, Soukas A, Salaspuro M. The effect of moderate drinking and abstinence on serum and urinary β -hexosaminidase levels. *Drug Alcohol Depend* 1990; **25**: 35–8
- 120 Wehr H, Czartoryska B, Gorska D, Matsumoto H. Serum β -hexosaminidase and α -mannosidase activities as markers of alcohol abuse. *Alcohol Clin Exp Res* 1991; **15**: 13–5
- 121 Nystrom M, Perasalo J, Salaspuro M. Serum β -hexosaminidase in young university students. *Alcohol Clin Exp Res* 1991; **15**: 877–80
- 122 Hultberg B, Isaksson A, Berglund M, Alling C. Increase and time-course variations in β -hexosaminidase isoenzyme B and carbohydrate-deficient transferrin in serum from alcoholics are similar. *Alcohol Clin Exp Res* 1995; **19**: 452–6
- 123 Pitkanen E, Kyllastinen M, Koivula T, Hormila P. β -N-Acetylglucosaminidase and β -glucuronidase activities in insulin-dependent diabetic subjects with retinopathy. *Diabetologia* 1980; **18**: 275–8
- 124 Hultberg B, Isaksson A, Lindgren A, Israelsson B, Brattstrom L. Plasma β -hexosaminidase isoenzymes A and B in patients with cerebral infarction. *Clin Chim Acta* 1996; **244**: 35–44
- 125 Hultberg B, Isaksson A. Isoenzyme pattern of serum β -hexosaminidase in liver disease, alcohol intoxication and pregnancy. *Enzyme* 1983; **30**: 166–71
- 126 Mahuran D, Novak A, Lowden JA. The lysosomal hexosaminidase isoenzymes. *Curr Top Biol Med Res* 1985; **12**: 229–88
- 127 DiPadova C, Worner TM, Lieber CS. Effect of abstinence on the blood acetaldehyde response to a test dose of alcohol in alcoholics. *Alcohol Clin Exp Res* 1987; **11**: 559–61
- 128 Hoffmann T, Meyer RJ, Sorrell MF, Tuma DJ. Reaction of acetaldehyde with proteins: formation of stable fluorescent adducts. *Alcohol Clin Exp Res* 1993; **17**: 69–74
- 129 Tuma DJ, Newman MR, Donohue TM, Sorrell MF. Covalent binding of acetaldehyde to proteins: participation of lysine residues. *Alcohol Clin Exp Res* 1987; **11**: 579–84
- 130 Stevens VJ, Fantl WJ, Newman CB, Sims RV, Cerami A, Peterson CM. Acetaldehyde adducts with haemoglobin. *J Clin Invest* 1981; **67**: 361–9
- 131 Hazelett SE, Liebelt RA, Truitt EB. Improved separation of acetaldehyde-induced hemoglobin. *Alcohol Clin Exp Res* 1993; **17**: 1107–11
- 132 Niemela O, Israel Y. Hemoglobin-acetaldehyde adducts in human alcohol abusers. *Lab Invest* 1992; **67**: 246–52
- 133 Niemela O, Juvonen T, Parkkila S. Immunohistochemical demonstration of acetaldehyde-modified epitopes in human liver after alcohol consumption. *J Clin Invest* 1991; **87**: 1367–74
- 134 Peterson CM, Polizzi CM. Improved method for acetaldehyde in plasma and haemoglobin-associated acetaldehyde: results in teetotalers and alcoholics reporting for treatment. *Alcoholism* 1987; **4**: 477–80
- 135 Peterson CM, Jovanovic-Peterson L, Schmid-Formby F. Rapid association of acetaldehyde with haemoglobin in human volunteers after low dose ethanol. *Alcoholism* 1988; **5**: 371–4
- 136 Hernandez-Munoz R, Ma X-L, Baraona E, Lieber CS. Method of acetaldehyde measurement with minimal artifactual formation in red blood cells and plasma of actively drinking subjects with alcoholism. *J Lab Clin Med* 1992; **120**: 35–41
- 137 Sillanaukee P, Seppa K, Koivula T. Effect of acetaldehyde on hemoglobin: HbA1-Ach as a potential marker of heavy drinking. *Alcohol* 1991; **8**: 377–81
- 138 Wickramasinghe SN, Corridan B, Hasan R, Marjot DH. Correlation between acetaldehyde-modified haemoglobin, carbohydrate-deficient transferrin (CDT) and haemoglobin abnormalities in chronic alcoholism. *Alcohol Alcohol* 1994; **29**: 415–23
- 139 Hazelett SE, Liebelt RA, Brown WJ, Androulakis V, Jarjoura D, Truitt EB Jr. Evaluation of acetaldehyde-modified hemoglobin and other markers of chronic heavy alcohol use: effects of gender and hemoglobin concentration. *Alcohol Clin Exp Res* 1998; **22**: 1813–9
- 140 Worrall S, de Jersey J, Shanley BC, Wilce PA. Antibodies to acetaldehyde-modified epitopes: an elevated immunoglobulin A response in alcoholics. *Eur J Clin Invest* 1991; **21**: 90–5
- 141 Worrall S, de Jersey J, Wilce PA, Seppa K, Hurme L, Sillanaukee P. Relationship between alcohol intake and immunoglobulin A immunoreactivity with acetaldehyde-modified bovine serum albumin. *Alcohol Clin Exp Res* 1996; **20**: 836–40
- 142 Sarkola T, Eriksson CJP, Niemela O, Sillanaukee P, Halmesmaki E. Mean cell volume and gamma glutamyl transferase are superior to carbohydrate-deficient transferrin and haemoglobin-acetaldehyde adducts in the follow-up of pregnant women with alcohol abuse. *Acta Obstet Gynecol Scand* 2000; **79**: 359–66

- 143 Doyle KM, Cluette-Brown JE, Dube DM, Bernhardt TG, Morse CR, Laposata M. Fatty acid ethyl esters in the blood as markers of ethanol intake. *JAMA* 1996; **276**: 1152–6
- 144 Hansson P, Caron M, Johnson G, Gustausson L, Alling C. Blood phosphatidylethanol as a marker of alcohol abuse: levels in alcoholic males during withdrawal. *Alcohol Clin Exp Res* 1997; **21**: 108–10
- 145 Sillanaukee P, Ponnio M, Seppa K. Sialic acid: new potential marker of alcohol abuse. *Alcohol Clin Exp Res* 1999; **23**: 1039–43
- 146 Agarwal DP, Tober-Rojas L, Harada S, Goedde HW. Comparative study of erythrocyte aldehyde dehydrogenase in alcoholics and control subjects. *Pharmacol Biochem Behav* 1983; **18**: 89–95
- 147 Shaw S, Stimmel B, Lieber CS. Plasma alpha amino-n-butyric acid to leucine ratio: an experimental marker of alcoholism. *Science* 1976; **194**: 1057–8
- 148 Collins MA, Nijm WP, Borge GF, Teas G, Goldfarb C. Dopamine-related tetrahydroisoquinolines: significant urinary excretion by alcoholics after alcohol consumption. *Science* 1979; **206**: 1184–6
- 149 Roine RP, Turpeinen U, Ylikahri R, Salaspuro M. Urinary dolcichol – a new marker of alcoholism. *Alcohol Clin Exp Res* 1987; **11**: 525–7
- 150 Anton RF, Moak DH. Carbohydrate-deficient transferrin and gamma-glutamyltransferase as markers of heavy alcohol consumption: gender differences. *Alcohol Clin Exp Res* 1994; **18**: 747–54
- 151 Helander A, Voltaire Carlsson A, Borg S. Longitudinal comparison of carbohydrate-deficient transferrin and gamma-glutamyltransferase complementary markers of excessive alcohol consumption. *Alcohol Alcohol* 1996; **31**: 101–7
- 152 Yersin B, Nicolet JF, Decrey H, Burnier M, van-Melle G, Pecoud A. Screening for excessive alcohol drinking. Comparative value of carbohydrate-deficient transferrin, gamma-glutamyltransferase and mean corpuscular volume. *Arch Intern Med* 1995; **155**: 1907–11
- 153 Stamm D, Hansert W, Feuerlein W. Excessive consumption of alcohol in men as a biological influence factor in clinical laboratory investigations. *J Clin Chem Clin Biochem* 1984; **22**: 65–7
- 154 Sillanaukee P. The diagnostic value of a discriminant score in the detection of alcohol abuse. *Arch Pathol Lab Med* 1992; **116**: 924–9
- 155 Rubio C, Gil V, Aparicio JM, Belda J, Pascual R, Merino J. Diagnostic efficiency of biological markers of alcohol consumption for the detection of excessive drinkers. *Ann Med Intern* 1996; **13**: 274–8
- 156 Weill J, Schellenberg F. The mathematics of measurement: sensitivity, specificity and predictive value of available tests. *Alcohol Alcohol Suppl* 1993; **2**: 107–10
- 157 Hillers VN, Alldredge JR, Massey LK. Determination of habitual alcohol intake from a panel of blood chemistries. *Alcohol Alcohol* 1986; **21**: 199–205
- 158 Rosalki SB. The clinical biochemistry of alcohol. In: Williams D, Marks V, eds. *Scientific Foundations of Biochemistry in Clinical Practice*, 2nd edn. Oxford: Butterworth-Heinemann, 1994

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